

3.7

0 🕗

0

DEVELOPMENT AND IMPLEMENTATION OF A CONCENTRATION AND DETECTION SYSTEM FOR INLINE MONITORING OF WATERBORNE PATHOGENS IN RAW AND DRINKING WATER

00

9

SPONSORED BY THE

θ

Federal Ministry of Education and Research

θ

0 0



0

RESULTS · APRIL 2017

## **PROJECT PROFILE**

## DEVELOPMENT AND IMPLEMENTATION OF A CONCENTRATION AND DETECTION SYSTEM FOR INLINE MONITORING OF WATERBORNE PATHOGENS IN RAW AND DRINKING WATER

2ND EDITION, APRIL 2017

Editors:	Dr. Daniel Karthe   Dr. Gregory Dame
Layout:	perner&schmidt werbung und design gmbh
Funding:	German Federal Ministry of Education and Research
Project Administration:	Project Management Jülich (PTJ)



# CONTENTS

1	INTRODUCTION	4
2	PROJECT CONSORTIUM	5
3	BACKGROUND	6
5		
4	TECHNICAL APPROACH AND WORKFLOW	8
5		Q
5		
6	DETECTION OF PATHOGENS	11
7		12
/		15
8	DATA MANAGEMENT	14
0	PROJECT RECUITE RURUEATIONS OUTLOOK	45
9		15
	CONTACT	16



## **1** INTRODUCTION

Laboratory for the surveillance of drinking water quality, Photo: André Künzelmann, UFZ The provision of hygienically safe drinking water was one of the greatest societal advances of the 20th century in the industrialized countries. In the early 21st century, however, ageing infrastructures as well as climate and demographic changes constitute new challenges for public water supply.

The project "Development and Implementation of a Concentration and Detection System for the Inline Monitoring of Waterborne Pathogens in Raw and Drinking Water" (EDIT) aimed at the development and pilot testing of a rapid detection system for bacteria and viruses that is suitable for application in the water industry. The system was designed to reduce the time needed for pathogen detection in raw and drinking water and to screen samples for more microbiological parameters from larger volumes than is possible with established methods. It is expected that in the future, there will be an increasing international demand for such systems – not only for drinking water surveillance but also for the monitoring of wastewater treatment or bathing water quality.

The EDIT project was supported by the German Federal Ministry of Education and Research in the context of the funding initiative "Smart and Multifunctional Infrastructural Systems for Sustainable Water Supply, Sanitation and Stormwater Management" (INIS) within the "Sustainable Water Management" (NaWaM) program.

# 2 PROJECT CONSORTIUM





Chair of Analytical Chemistry and Institute of Hydrochemistry (IWC), Technical University of Munich

Institute for Microsystems Technology, University of Freiburg



Technologiezentrum Wasser (TZW, Technology Center for Water) of the German Technical and Scientific Association for Gas and Water, Karlsruhe



HELMHOLTZ ZENTRUM FÜR UMWELTFORSCHUNG UFZ







Application Center for Systems Technology, Fraunhofer Institute of Optronics, System Technologies and Image Exploitation (IOSB), Ilmenau

Helmholtz Centre for Environmental Research GmbH – UFZ, Magdeburg

Institute of Microbiology and Virology, Brandenburg Medical School Theodor Fontane, Senftenberg

Municipal Water Supply Company of Berlin

GWK Präzisionstechnologie GmbH, Munich

R-Biopharm AG, Darmstadt

## **ASSOCIATED PARTNERS:**

Municipal water supply company of Magdeburg – TWM | Municipal water supply company of Marburg – SWM Department of Geography, Georg-August University, Göttingen



Preparing samples for water quality surveillance, Photo: André Künzelmann, UFZ

## 3 BACKGROUND



The drinking water supply in Germany is considered to be of very high standard. Supply shortfalls and impairments of drinking water quality occur only in exceptional situations. However, the relatively old age of supply infrastructures, climate change and demographic change result in new challenges which are relevant for public water supply.

## IMPACTS OF CLIMATE CHANGE

Observed and expected climate changes in Germany have significant but spatially heterogeneous impacts on regional hydrology and water availability. A predicted increase in the frequency and severity of hydro-meteorological extremes (i.e. floods and droughts) is likely to lead to reduced raw water quality and potentially even the contamination of drinking water supply and distribution systems. Moreover, increasing water temperatures can also favor the survival and reproduction of hygienically relevant microorganisms. The challenges resulting from climate change tend to be aggravated by demographic changes.

### **IMPACTS OF DEMOGRAPHIC CHANGE**

In the aftermath of Germany's reunification, some parts of eastern Germany experienced massive population shrinkage. Low birthrates and internal migration towards dynamically developing urban regions continue to lead to a depopulation of smaller cities and peripheral rural areas. The reduction in water demand in affected regions is exacerbated by a declining per capita consumption, with negative impacts on the degree of capacity utilization in water works and distribution systems. Without countermeasures, the resulting increase in water transit times in pipelines and intermediate storage tanks may also promote the growth of harmful microorganisms.

Samples for microbiological examination, Photo: André Künzelmann, UFZ



#### Relative frequency that hygienic norms were exceeded



The map shows the total number of cases that any hygienic parameter exceeded the maximum limit per 10,000 people. Only such water supply areas are shown that have more than 5000 inhabitants or a daily supply of more than 1000 m<sup>3</sup>

Cartography: Niklas Rehkopp (2015) Source: Federal Agency for Carthography and Geodesy (2013) Federal Ministry of Health and Federal Enviroment Agency (2011, 2015)

			Kilometers
D 10	20 20	00	400

## NEED FOR INNOVATIVE SURVEILLANCE METHODS

In the light of these challenges, rapid and reliable methods for the surveillance of drinking water hygiene could help to ensure a safe water supply in the future. Currently used methods are based on the cultivation of indicator bacteria. The main disadvantage is the time needed for detection and quantification of these bacteria (typically at least 18 hours). Moreover, even negative test results cannot completely rule out a contamination with different micro-organisms, especially viruses.

Time and labor-intensive cultivation techniques are still the standard for surveillance of water hygiene, Photo: Dr. Daniel Karthe, UFZ





Flowchart of the hygiene online monintoring, Source: EDIT Consortium

 CUF:
 Crossflow ultrafiltration

 MAF:
 Monolithic adsorption filtration

 ceUF:
 Centrifugal ultrafiltration

 isoAmp:
 isothermal amplification

 MCR3:
 Microarray chip reader 3

 SIMS:
 Systemically integrated management system

 OPC:
 OLE for Process Control (standardized software interface)

## 4 TECHNICAL APPROACH AND WORKFLOW





Upper photo: Development of the lab-on-chip system Lower photo: Demonstration of the CUF system to end users at BWB, Photos: Dr. Daniel Karthe, UFZ

The EDIT project aimed at the development of an online monitoring system for hygiene which allows for the continuous surveillance and rapid detection of harmful microorganisms in raw and drinking water.

As compared to state-of-the-art techniques in which a small sample volume (100 ml) is placed on selective cultivation media, the EDIT approach is based on the combination of multiple concentration steps and the subsequent detection of bacteria and viruses using molecular biology.

## FILTRATION, PREPARATION AND DETECTION STEPS:

- concentration of samples in four steps from several hundred/ thousand liters to about 10 µl;
- sample purification and extraction of microorganisms;
- module for discrimination between active and inactive microorganisms and viruses (in order to mark harmless microorganisms such as those inactivated by sterilization);
- detection of (harmful) microorganisms and viruses by molecular methods (based on DNA or RNA);
- data interpretation and communication of results.



CUF and MAF concentration systems being tested at TUM, Photo: PD Dr. Michael Seidel, TUM

## 5 CONCENTRATION AND EXTRACTION OF MICROORGANISMS

The fact that exposure to even a small number of pathogenic microorganisms or viruses can have detrimental health effects is a major challenge for the surveillance of drinking water. In order to permit a reliable risk assessment, water samples of a sufficient volume need to be analyzed. Because state-of-the-art detection systems cannot handle very large volumes, the EDIT project established a method to concentrate samples from several hundred to several thousand liters to a few milliliters (macro-concentration) and subsequently reach a volume of a few microliters in a lab-on-chip system (micro-concentration). The lab-on-chip system also allows for the extraction of DNA or RNA of target pathogens, which can then be amplified and detected on the microarray analysis system MCR 3.

## MACROCONCENTRATION OF BACTERIA AND VIRUSES

Jointly with GWK Precision Technology, the Institute of Hydrochemistry at Technical University of Munich developed several sequential concentration modules for raw and drinking water. In a first step, bacteria and viruses are concentrated via crossflow ultrafiltration (CUF) at a rate of about 1000 l/h from several hundred to thousand liters of raw or drinking water to about 20 l.

Development and production of the lab-on-chip system, Photos: Dr. Daniel Karthe, UFZ









Lab-on-chip system, Photo: Matthias Hügle, MHB

The next step consists of a large system for monolithic adsorption filtration (BigMAF), which concentrates bacteria and viruses from 20 I samples to about 20 ml, thereby removing most of the matrix components. This is followed by a centrifugal ultrafiltration (ceUF), which further reduces the sample volume to about 1 ml. The time needed for these two steps is about 45 min.

## LAB-ON-CHIP MICROCONCENTRATION AND EXTRACTION OF NUCLEIC ACIDS

The subsequent micro-concentration and extraction of nucleic acids from the macro-concentrated sample were implemented on a single lab-on-chip system. In this system, suspended bacteria and viruses are concentrated at a specific gel front with high efficiency using the principle of free flow electrophoresis. The volume in the chip is reduced to about 10 µl after accumulation of microorganisms. Lysis of microorganisms and the extraction of nucleic acids are performed in the same lab-on-chip system. In the next step the released nucleic acids are purified via gel electrophoresis. After this the extract is transmitted to the automated analytical microarray (MCR3).

0

Chip for micro-concentration and sample extraction, Photo: Richard Rietzel, IMTEK Freiburg

DEVELOPMENT AND IMPLEMENTATION OF A CONCENTRATION AND DETECTION SYSTEM FOR



DNA Microarray System, Photo: GWK & TUM

## 6 DETECTION OF PATHOGENS

Currently used methods for the assessment of drinking water hygiene are based on the cultivation of indicator bacteria in the laboratory. By contrast, the hygiene online monitoring (HOLM) system developed in EDIT is based on molecular biology. Due to the multiplex amplification and detection approach, several indicators and pathogens can be identified simultaneously and within much shorter time than conventional methods (about 5 hours instead of 18 hours).

In the project, two methods for nucleic acid amplification systems were considered: polymerase chain reaction (PCR) and an isothermal amplification technique (Recombinase Polymerase Assay, RPA). While applications for RPA in medical diagnostics have increased significantly in recent years, the technique has the additional advantage of combining a high amplification (and therefore detection) speed with relatively inexpensive and less complex technology as compared to PCR. This facilitated the integration into the HOLM system. Despite advances, the simultaneous amplification of different DNA sequences (i.e. multiplexing) that are useful for the identification of waterborne bacteria and viruses remains challenging.

For the detection of the specific DNA of pathogens and indicators, a DNA microarray was designed for the HOLM system. With species specific DNA oligomers generated by isothermal amplification the pathogen nucleic acids are detected *in situ* via chemiluminiscence on the analytical microarray, allowing for the direct detection of multiple pathogenic microorganisms and viruses.



Pathogen detection on the MCR3, Source: Dr. Andreas Kunze, TUM

## MICROORGANISMS CONSIDERED IN EDIT

Based on the simultaneous detection of several microorganisms, the HOLM system developed in EDIT can identify a wide range of bacteria, viruses and phages (see table 1) that exceeds the requirements of the German Drinking Water Ordinance. Viruses are particularly relevant in this context because of the large amounts excreted in feces, low infective doses and the high environmental stability of viruses may promote their occurrence in raw water used for drinking water or food production.

Bacteria	Viruses	Bacteriophages	
• Escherichia coli <sup>1</sup>	Norovirus GI, II	• MS2	1 -
• Enterococcus faecalis <sup>1</sup>	• Adenovirus 40, 41, 52	• PhiX174	®
• Pseudomonas aeruginosa <sup>1</sup>	Enteroviruses		
• Legionella pneumophila <sup>1</sup>			-
Campylobacter jejuni			
• Klebsiella pneumoniae <sup>2</sup>			18
and Klebsiella oxytoca			

Table 1: Target organisms of the amplification and detection module 1Regulated by the German Drinking Water Ordinance, <sup>2</sup>Coliform bacteria



Color marking of inactive microorganisms, Source: Dr. Johannes Ho, TZW

## DISCRIMINATION BETWEEN ACTIVE AND INACTIVE MICROORGANISMS

Drinking water treatment sometimes relies on physical or chemical disinfection methods to inactivate microorganisms and thus ensure hygienic water safety. Such treatments can damage microorganisms in various ways. For instance, bacteria or viruses can no longer replicate or infect a person after their membranes or capsids are damaged. However, the genome may be still undamaged and can thus be detected by molecular methods. The (positive) detection of bacteria or viruses which are no longer infectious would constitute an undesirable, false-positive result for drinking water surveillance.



microorganisms, Photo: Dr. Johannes Ho, TZW

MARKING DAMAGED CELLS BY PROPIDIUM MONOAZIDE (PMA)

their detection by molecular methods. By contrast, PMA cannot penetrate undamaged cells and viruses. In this way, only active microorganisms are detected while inactive organisms are excluded. The left picture shows a water sample with dead (red, PMA) and living (green, SYBR Green) cells.

Due to their complex structure, bacteria and viruses can be damaged in different ways by disinfection. Depending on the detection method, different characteristics are used: reproduction (culture methods), genomes (PCR and RPA), membrane or capsid damages (PMA-PCR), amount of DNA and size of cells (flow cytometry). Different methods differ in their test results when cells or virus particles are damaged. The definitions for live and dead (active or inactive) microorganisms are, however, not unambiguous and currently the subject of scientific discussion. In the EDIT project, different detection methods were compared to increase the information about microorganisms exposed to different disinfection methods.

### INTEGRATION INTO THE HOLM SYSTEM

The treatment with PMA is implemented after the concentration of water samples and prior to the sample preparation for amplification and detection. The application of PMA was therefore integrated into the lab-on-chip system.

## PMA is adsorbed to the nucleic acids of damaged cells or virus particles and in this way inhibits

kontin Crossillow Libra-Fi	Righter.	549	
Menge Eingang (I) 80	.00		
Menge Abgabe     70	.00		
Editratifices (U/h) 250	0,00		
Eingungsdruck (tur) . 1,	56		
Verhältnis Probe zu Gesam 1.	ethus (1:s) 05		
Trammembrandruck minim 0,	uil (tar) 80		
Tranumentbrandruck mails 3,	nat (bar) 00		
Transmensbrandruck final 1,	11-1 22		
Genti.			



Right picture: Automatic position marking during sampling

Left picture: Data entry during sample concentration, Source: Dipl.-Ing. Thomas Westerhoff, Fraunhofer AST

## 8 DATA MANAGEMENT

The hygiene online monitoring system (HOLM) developed in EDIT consists of several modules for unique steps of sample processing. These subsystems had to be connected to each other not only for the physical transfer of samples, but also in a way that for the complete workflow of any sample all results and operational parameters are documented. This is not only a prerequisite for quality-assurance, but also for system-internal plausibility and validity checks.

### DATA CAPTURE

For the capture, storage and analysis of data at different process steps, the Fraunhofer AST created an online database using PostgreSQL. Data can be entered both automatically and manually via an app for mobile devices. For security reasons, all data are transported in an encrypted form. Individual samples can be identified by scanning QR code on sample containers with a smartphone camera. Location data can automatically added by the smartphone's GPS functionality. For the direct and automated transmission of analytical data and operational parameters to mobile devices, the implementation of interfaces such as NFC or Bluetooth is planned for the future. Results obtained on the analytical microarray are automatically evaluated. When relevant microoranisms are detected, an alarm is automatically transmitted to the waterworks' control system via a standardized OPC interface. A subsequent software-based simulation of the spread of pathogens in the distribution network can further help to plan intervention strategies.

### INTELLIGENT DATA MANAGEMENT

Seamless communication from sensors to an online database used by end users is a good example for Industry 4.0, which is an important element of the high-tech strategy of the German Federal government which aims at the creation of added value by intelligent application of IT in industry.



Demonstration of the lab-on-chip system and MCR 3 at TUM Photo: Dr. Daniel Karthe, UFZ

# 9 PROJECT RESULTS, PUBLICATIONS, OUTLOOK

In the EDIT project, a modular hygiene online monitoring system was developed and tested both under laboratory conditions and by future end users in the water industry. The feasibility and potential of combining multiple concentration steps, further sample pretreatment and DNA extraction on a lab-on-chip system, and RPA-based amplification and detection on an analytical microarray could be demonstrated. However, future research is needed to validate the technique under real-life and laboratory conditions and to further improve the system components in order to develop a prototype that is cost-effective and widely acceptable by end-users.

## PUBLICATIONS

HAKENBERG, S.; HÜGLE, M.; MEYER, P.; BEHRMANN, O.; DAME, G. & URBAN, G.A. (2015): Fenton fragmentation for faster electrophoretic on chip purification of amplifiable genomic DNA. Biosensors and Bioelectronics 67:59-52. doi: 10.1016/j.bios.2014.06.003

HO, J.; SEIDEL, M.; NIESSNER, R.; EGGERS, J. & TIEHM, A. (2016): Long amplicon (LA)-qPCR for the discrimination of infectious and noninfectious phix174 bacteriophages after UV inactivation. Water Research 103:141-148. doi: 10.1016/j.watres.2016.07.032

KARTHE, D. (2015): Bedeutung hydrometeorologischer Extremereignisse im Kontext des Klimawandels für die Trinkwasserhygiene in Deutschland und Mitteleuropa. Hydrologie und Wasserbewirtschaftung 59(5):264-270. doi: 10.5675/HyWa\_2015,5\_7 KARTHE, D.; BEHRMANN, O.; BLÄTTEL, V.; ELSÄSSER, D.; HEESE, C.; HO, J.; HÜGLE, M.; HUFERT, F.; KUNZE, A.; NIESSNER, R.; SCHARAW, B.; SPOO, M.; TIEHM, A.; URBAN, G.; VOSSELER, S.; WESTERHOFF, T.; DAME, G. & SEIDEL, M.: Modular Development of an Inline Monitoring System for Waterborne Pathogens in Raw and Drinking Water. Environmental Earth Sciences 75:1481. doi: 10.1007/s12665-016-6287-9

KARTHE, D.; REHKOPP, N.; REEH, T. & FAUST, H. (2016): Regional Disparities of Microbiological Drinking Water Quality: Assessment of Spatial Pattern and Potential Sociodemographic Determinants. Urban Water Journal. doi: 10.1080/1573062X.2016.1240809

KUNZE, A.; DILCHER, M.; ABD EL WAHED, A.; HUFERT, F.; NIESSNER, R. & SEIDEL, M. (2016): On-Chip isothermal nucleic acid amplification on flow-based chemiluminescence microarray analysis platform for the detection of viruses and bacteria. Analytical Chemistry 88(1):898–905. doi: 10.1021/acs.analchem.5b03540

KUNZE, A.; PEI, L.; ELSÄSSER, D.; NIESSNER, R. & SEIDEL, M. (2015): High performance concentration method for viruses in drinking water. Journal of Virological Methods 222C:132-137. doi:10.1016/j. jviromet.2015.06.007

LENGGER, S.; OTTO, J.; ELSÄSSER, D.; SCHNEIDER, O.; TIEHM, A.; FLEISCHER, J.; NIESSNER, R. & SEIDEL, M. (2014): Oligonucleotide microarray chip for the quantification of MS2, PhiX174, and adenoviruses on the multiplex analysis platform MCR 3. Analytical and Bioanalytical Chemistry 14:3323-3334. doi: 10.1007%2Fs00216-014-7641-y.

## OUTLOOK

A need for rapid and automated systems for the surveillance of pathogens in water exists not only in Germany but worldwide. However, future research is still needed to come to a prototype that is costeffective and widely acceptable by end-users. In particular, the following aspects need to be addressed:

- validation of the technique under real-life and laboratory conditions;
- improvement of the system components and complete automation of the system;
- optimization of size and handling of the system according to enduser requirements;
- minimization of maintenance and operational costs

Ultimately, such systems have a potential for the hygienic monitoring not only of raw and drinking water, but also in specific water cycles (e.g. in large buildings or on cruise ships), in the food industry or for bathing waters.

## CONTACT

#### Helmholtz-Centre for Environmental Research GmbH – UFZ

Department Aquatic Ecosystem Analysis and Management Brückstraße 3a | 39114 Magdeburg | Dr. Daniel Karthe | daniel.karthe@ufz.de

### Brandenburg Medical School Theodor Fontane, Senftenberg

Institute of Microbiology and Virology Universitätsplatz 1 | 01968 Senftenberg | Dr. Gregory Dame/Prof. Dr. Frank Hufert | dame@mhb-fontane.de

### **Technical University of Munich**

Chair of Analytical Chemistry and Institute of Hydrochemistry Marchioninistraße 17 | 81377 München | PD Dr. Michael Seidel/Prof. Dr. Reinhard Niessner | michael.seidel@ch.tum.de

## German Technical and Scientific Association for Gas and Water - DVGW

Technologiezentrum Wasser (TZW) Karlsruher Straße 84 | 76139 Karlsruhe | Prof. Dr. Andreas Tiehm | andreas.tiehm@tzw.de

#### Fraunhofer Institute of Optronics, System Technologies and Image Exploitation – IOSB

Application Center for Systems Technology (AST) Am Vogelherd 50 | 98693 Ilmenau | Dr.-Ing. Buren Scharaw | buren.scharaw@iosb-ast.fraunhofer.de

### Albert-Ludwigs-University, Freiburg

Institute of Microsystems Technology (IMTEK) Georges-Köhler-Allee 103 | 79110 Freiburg | Prof. Dr. Gerald Urban | urban@imtek.de

## Municipal Water Supply Company of Berlin (Berliner Wasserbetriebe) - BWB

Neue Jüdenstraße 1 | 10179 Berlin | Dipl.-Ing. Fereshte Sedehizade | fereshte.sedehizade@bwb.de

### **GWK Präzisionstechnik GmbH**

Gollierstraße 70 | 80339 München | Christian Heese | christian.heese@gwk-munich.com

### **R-Biopharm AG**

An der neuen Bergstraße 17 | 64297 Darmstadt | Dr. Silvia Vosseler | s.vosseler@r-biopharm.de





https://nawam-inis.de/en/inis-projects/edit